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## Suspension of Fe<sub>3</sub>O<sub>4</sub> nanoparticles stabilized by chitosan and *o*-carboxymethylchitosan

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### Abstract

In this study, a well-dispersed suspension of superparamagnetic Fe<sub>3</sub>O<sub>4</sub> nanoparticles was stabilized by chitosan (CS) and *o*-carboxymethylchitosan (OCMCS), respectively. The resulting magnetic Fe<sub>3</sub>O<sub>4</sub> nanoparticles were characterized by dynamic light scattering (DLS), Fourier transform infrared spectroscopy (FTIR), X-ray diffraction (XRD), transmission electron microscope (TEM), zeta-potential measurement and vibrating sample magnetometry (VSM). TEM results demonstrated a spherical or ellipsoidal morphology with an average diameter of 14–20 nm. The adsorbed layer of CS and OCMCS on the magnetic surface was confirmed by FTIR. XRD illustrated that the resulting magnetic nanoparticles have a spinel structure and lastly VSM results showed the modified magnetic Fe<sub>3</sub>O<sub>4</sub> nanoparticles were superparamagnetic. The adsorption mechanism of CS and OCMCS onto the surface of Fe<sub>3</sub>O<sub>4</sub> nanoparticles is believed to be the electrostatic and coordination interactions, respectively. The mechanisms of both CS and OCMCS stabilizing the suspension of Fe<sub>3</sub>O<sub>4</sub> nanoparticles were supposed electrostatic repulsion. These well-dispersed superparamagnetic Fe<sub>3</sub>O<sub>4</sub> nanoparticles stabilized by the biocompatible CS or OCMCS dispersant should have potential applications in biotechnology fields. © 2007 Elsevier B.V. All rights reserved.

Keywords: Superparamagnetic; Fe<sub>3</sub>O<sub>4</sub>; Nanoparticles; Well-dispersed; Chitosan; o-Carboxymethylchitosan

### 1. Introduction

The synthesis of uniformly sized magnetic nanoparticles has been intensively pursued because of their broad applications, including magnetic storage media, ferrofluids, magnetic resonance imaging, and magnetically guided drug delivery (Kim et al., 2001a,b; Reynolds et al., 2000; Lubbe et al., 1996; Chan et al., 1993; Jordan et al., 1999; Dyal et al., 2003; Neuberger et al., 2005; Morales et al., 2005). Various common protocols were developed to synthesize iron oxides, such as condensing divalent and trivalent iron salts in reactions with hydroxide bases (pH 9.5–10) (Mann and Hannington, 1988; Kim et al., 2001a,b; Zhang et al., 2002; Harris et al., 2003), and chemical oxidation in micelle media (Shen et al., 1999) or in polymer (Ma et al., 2005; Xie et al., 2004).

However, one of the main challenges facing almost all of these novel techniques is the ability to transport and deliver well-

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dispersed nanoparticles with the desired composition, structure, and uniformity as well as the prevention of aggregation (Chouly et al., 1996; Thode et al., 1997; Mahapatra et al., 2006). A crucial physical property of magnetic nanoparticles is their tendency to aggregate. Encounters between particles dispersed in liquid media occur frequently, and the stability of a suspension is determined by the interaction between the particles during these encounters. The principal cause of aggregation is the short-range forces—van der Waals attraction between the two particles. To counteract these attractive interactions and promote stability, equally short-range repulsive forces are required (Bonnemann et al., 2001). They are enforced either by electrostatic repulsion between the particles or by coating the particles with organic long-chain molecules (Chang et al., 2006; Zhi et al., 2006).

Magnetic nanoparticles can be dispersed in carrier fluid through specific interactions between the particle surfaces and selected low-molecular-weight surfactants or polymer. Various methods have been reported for the preparation of stable dispersions of iron oxide in organic solvents (organosols), including hexane and decane (Shafi et al., 2001; Fried et

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al., 2001). However, for these organosols, the biological applications are greatly restricted because of their poor solubility in aqueous solutions. For biomedical applications, it is essential to develop nanoscale magnetite with a narrow size distribution and surface coating with biocompatibility materials, i.e., nonimmunogenic, nonantigenic, and protein-resistant (Xie et al., 2004; Chouly et al., 1996; Shieh et al., 2005).

Due to its biocompatibility and biodegradability, chitosan has been widely employed in tissue engineering and controlled drug/gene delivery (Kofuji et al., 2001; Thanou et al., 2001; Roy et al., 1999). Chitosan with net positive charge binds with recombinant DNA plasmids to form complex nanoparticles for delivering genes into cells (Mao et al., 2001; Artursson et al., 1994; Senel et al., 2000; Fang et al., 2001). The bio-adhesiveness of chitosan makes it an ideal candidate for mucosal drug delivery (Fang and Chan, 2003). For instance, the co-administration of chitosan with drugs has been shown to enhance the transcellular and paracellular transport of drugs across mucosal epithelium (Chan et al., 2001). Moreover, the overcoming of the cellular membrane barrier by chitosan-based gene carriers in the absence of receptor-mediated endocytosis has led to the hypothesis that chitosan directly interacts with cell membranes (Thanou et al., 2001). o-Carboxymethylchitosan (OCMCS) has a backbone structure similar to CS, but the o-hydroxyl group of each monomer is substituted by a carboxymethyl group through ether bond formation (Scheme 1). OCMCS has been shown to have amphiphilic, blood compatible and effective membrane penetrable properties (Zhu et al., 2005a,b). More strikingly, it can load hydrophobic anticancer drugs effectively (Zhu et al., 2006). Though lots of synthesized polymers (e.g., poly(vinyl alcohol) phosphate, polyethylene glycol, polyamides, polyglycidyl methacrylate, poly(acrylic acid)) were employed as a coating agent in the surface modification of iron oxide particles (Gas et al., 2006; Guo et al., 2007; Abu-Much et al., 2006; Wan et al., 2007; Xie et al., 2004), the natural polymeric and bioactive properties of chitosan and OCMCS lead our interest to explore the feasibility of synthesizing a well-dispersed aqueous dispersion of superparamagnetic Fe<sub>3</sub>O<sub>4</sub> nanoparticles stabilized by CS or OCMCS, simply and effectively. The mechanisms of CS and OCMCS stabilizing the suspension of  $Fe_3O_4$  nanoparticles were discussed in this study.

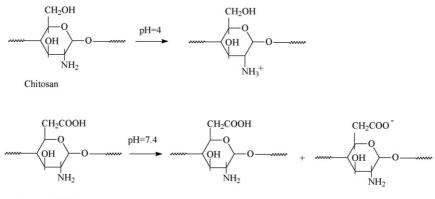
### 2. Experimental

### 2.1. Chemicals and materials

Ferric chloride hexahydrate (FeCl<sub>3</sub>·6H<sub>2</sub>O, >99%), ferrous chloride tetrahydrate (FeCl<sub>2</sub>·4H<sub>2</sub>O, >99%) and ammonium hydroxide (29.4%) were obtained from Shanghai Chemical Reagent (China). Chitosan was obtained from Lianyungang Biologicals Inc. (China). Its viscosity average molecular weight was  $5.2 \times 10^5$  g/mol, and the degree of deacetylation was 90%. *o*-Carboxymethylchitosan, containing about 100 carboxyl groups and 75 amino groups per 100 anhydroglucosamine units of chitosan was prepared by methods we have previously reported (Zhu et al., 2005a,b). All the other chemicals were of reagent grade and used without further purification.

### 2.2. Preparation of magnetic $Fe_3O_4$ nanoparticles

Fe<sub>3</sub>O<sub>4</sub> nanoparticles were prepared by the coprecipitation of ferric and ferrous salts in anaerobic conditions at ambient temperature. Fifty milliliters of 1.5 M ammonium hydroxide solution was put into a three-neck, round-bottom flask with magnetic stirrer with N2 protection. To this, 5 mL of iron solution containing 0.1 M Fe<sup>2+</sup> and 0.2 M Fe<sup>3+</sup> in double-distilled water was added slowly by a drop funnel, the color of the suspension turned black immediately, indicating the formation of magnetite. The resulting black precipitate was collected with a strong magnet, and the supernatant was removed from the precipitate by decantation. Deoxygenated double-distilled water was added to wash the powder. The solution was centrifuged for 2 min at a speed of 1800 rpm and the solution was decanted. Centrifugation-redispersion cycles were carried for several times to remove excess ammonia from the remaining solution. Finally, a black precipitate (magnetite) was obtained by freeze-drying.



o-carboxymethylchitosan

Scheme 1. Charge formation of chitosan and OCMCS in aqueous solution.

# 2.3. Chitosan and OCMCS surface-modified Fe<sub>3</sub>O<sub>4</sub> nanoparticles

For chitosan or OCMCS surface-modified  $Fe_3O_4$  nanoparticles, 10 mg as-prepared black precipitate was dispersed in 10 mL of 0.2 mg/mL of an acidic chitosan solution (pH 4) or 10 mL of 0.2 mg/mL OCMCS neutral aqueous solution (pH 7.4), respectively, and stirred for 12 h at room temperature to ensure the particles were coated evenly. The resulted black dispersions were purified by several centrifugation–redispersion cycles to remove the free CS and OCMCS. The colloidal solution of CS/Fe<sub>3</sub>O<sub>4</sub> or OCMCS/Fe<sub>3</sub>O<sub>4</sub> magnetic nanoparticles was centrifuged for 10 min at a speed of 25,000 rpm. Finally, CS/Fe<sub>3</sub>O<sub>4</sub> and OCMCS/Fe<sub>3</sub>O<sub>4</sub> magnetic nanoparticles were obtained by freeze-drying.

#### 2.4. Characterization

The crystal structure of the samples was examined by X-ray diffraction (XRD) with a XD-3A powder diffractometer, using a monochromatized X-ray beam with nickel-filtered Cu Ka radiation in the range of 5–40° (2 $\theta$ ) at 40 kV and 30 mA. The size and morphology of the magnetite nanoparticles were measured by a TE CHAI-12 (Philips) transmission electron microscopy (TEM). The particle size distribution was investigated by using a modified commercial LLS spectrometer (ALV/SP-125, Germany), which was equipped with a solid-state laser (Coherent DPSS) having an output power of 400 mW at  $\lambda_0$  532.0 nm and an ALV-5000 multi- $\tau$  digital time correlator, to perform the dynamic light scattering experiments. Fourier transform infrared (FTIR) spectroscopy was applied to characterize the changes in the chemical structure of Fe<sub>3</sub>O<sub>4</sub> nanoparticles after surface stabilization. The magnetization and hysteresis loop were measured at the room temperature with a Lake Shore Model 7300 VSM. The zeta potential of the magnetite suspension was found using a zeta-potential analyzer (Zetaplus, Brookhaven Instruments, USA).

### 3. Results and discussion

# 3.1. Stability, particle size and its distribution of colloidal solution

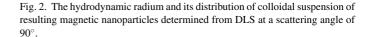
Both CS and OCMCS are supposed to be good surfaceactive agents for stabilizing aqueous suspension of  $Fe_3O_4$ magnetic nanoparticles because of their polyelectrolyte property. The colloidal stability of resulting magnetic nanoparticles is shown in Fig. 1, which clearly demonstrates that a CS or OCMCS stabilized magnetic nanoparticles exhibits a welldispersed appearance, while the unmodified dispersion of magnetic nanoparticles precipitates completely.

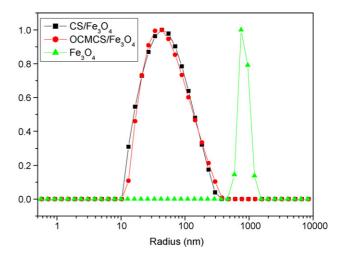
The particle size and its distribution of the suspension of unmodified, CS and OCMCS stabilized magnetic nanoparticles are shown in Fig. 2. The mean particle radius was measured by DLS is 42 nm with a polydispersity index of 0.262 for CS/Fe<sub>3</sub>O<sub>4</sub> nanoparticles, and 38 nm with a polydispersity index of 0.285 for OCMCS/Fe<sub>3</sub>O<sub>4</sub> nanoparticles. Though the high polydis-

Fig. 1. Stability comparison of colloidal suspension of CS, OCMCS and unmodified magnetic nanoparticles (from left to right) (all the suspensions were centrifuged at 10,000 rpm for 10 min).

persity index suggests that there is still some aggregation of CS and OCMCS stabilized  $Fe_3O_4$  nanoparticles in their suspension, the size of the hydrodynamic radium of CS/Fe<sub>3</sub>O<sub>4</sub> or OCMCS/Fe<sub>3</sub>O<sub>4</sub> particles is within the nanoscale range. However, the mean particle radius for unmodified magnetic nanoparticles shows 748 nm, suggesting a very fast and strong flocculation.

Colloidal processing of magnetic nanoparticles has been extensively studied with emphasis on the characteristics of the dispersion as a ferrofluid. Suspensions of magnetic nanoparticles contain van der Waals' forces and magnetic dipole–dipole interactions generated from residual magnetic moments, which tend to agglomerate and flocculate the particles. Ferrofluids composed of magnetic nanoparticles are not only affected by an inhomogeneous particle size distribution, but also by the surface charge of the particles in the solution. A practical method used to stabilize ferrofluids is through electrostatic repulsion, achieved by similar surface charge, which results from the formation of an electrical double layer around the particles. A







colloidal solution is a suspension in which the dispersed phase is so small that gravitational forces are negligible and particle interactions are dominated by short-range forces, such as van der Waals' attraction and surface charges. The inertia of the dispersed phase is small enough that it exhibits Brownian motion, a random walk driven by momentum transferred through collisions with molecules of the suspending medium (Khalafalla and Reimers, 1973). As above, the magnetic nanoparticles are usually required to graft with nonmagnetic substances to form a stable ferrofluid. Many kinds of natural (e.g., proteins and polysaccharides), synthetic (e.g., polyelectrolytes), and non-ionic polymers (e.g., poly(vinyl alcohol)) have been used for particle coating. The present results expressed that both chitosan and OCMCS are effective to disperse the magnetic Fe<sub>3</sub>O<sub>4</sub> nanoparticles. Moreover, the suspension of CS/Fe<sub>3</sub>O<sub>4</sub> and OCMCS/Fe<sub>3</sub>O<sub>4</sub> nanoparticles can maintain their initial properties more than 6 months, which is very important for its biomedical applications.

### 3.2. FTIR

The FTIR spectra of unmodified, CS and OCMCS surfacemodified magnetic nanoparticles are shown in Fig. 3. Fig. 3(a) shows the peak of 569 cm<sup>-1</sup>, which is typical characteristic of Fe–O–Fe in Fe<sub>3</sub>O<sub>4</sub>. However, from Fig. 3(b) and (c), it can be seen that the characteristic peak of Fe–O–Fe shifts to 575 and 583 cm<sup>-1</sup>, respectively. In addition, the peaks of 1580 and 1425 cm<sup>-1</sup> appeared in Fig. 3(b) are characteristic absorption bands of chitosan, and the peaks of 1558 and 1406 cm<sup>-1</sup> bands presented in Fig. 3(c) are characteristic absorption bands of OCMCS. These results proved that CS and OCMCS have been adsorbed onto Fe<sub>3</sub>O<sub>4</sub> nanoparticles.

### 3.3. TEM morphology

TEM was applied to determine the morphology of the magnetite and the results are shown in Fig. 4. It can be seen from Fig. 4 that the resulting magnetic  $Fe_3O_4$  nanoparticles are almost spherical or ellipsoidal. The mean particle size is 14.1 nm with a

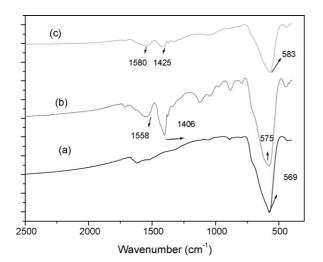


Fig. 3. FTIR spectra of unmodified, CS and OCMCS magnetic nanoparticles.

standard deviation of about 3.5 nm. The average particle size and standard deviation are 16.7 and 3.2 for CS/Fe<sub>3</sub>O<sub>4</sub> nanoparticles, respectively. The average particle size and standard deviation are 19.4 and 4.6 nm for OCMCS/Fe<sub>3</sub>O<sub>4</sub> nanoparticles, respectively. Though it is difficult to directly observe the adlayer of CS or OCMCS on the Fe<sub>3</sub>O<sub>4</sub> nanoparticles from TEM micro-

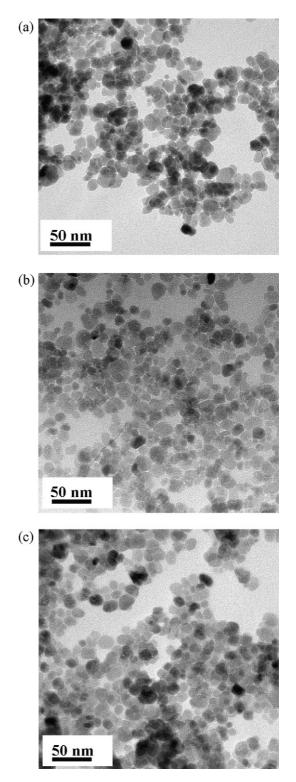


Fig. 4. TEM morphology of (a) unmodified  $Fe_3O_4$  magnetic nanoparticles; (b) CS/Fe<sub>3</sub>O<sub>4</sub> magnetic nanoparticles; (c) OCMCS/Fe<sub>3</sub>O<sub>4</sub> magnetic nanoparticles.

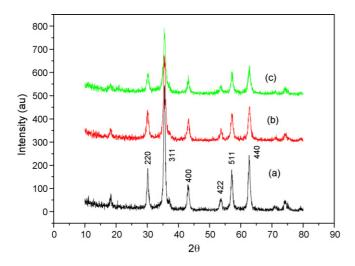


Fig. 5. X-ray powder diffraction patterns of the Fe<sub>3</sub>O<sub>4</sub> magnetic nanoparticles.

graphs, the dispersing behavior of surface stabilized  $Fe_3O_4$  nanoparticles (Fig. 1(b) and (c)) has obviously been improved in comparison with that of unmodified magnetic nanoparticles (Fig. 1(a)), which exhibits aggregated morphology.

### 3.4. XRD

To confirm the presence of crystalline Fe<sub>3</sub>O<sub>4</sub>, CS/Fe<sub>3</sub>O<sub>4</sub> and OCMCS/Fe<sub>3</sub>O<sub>4</sub> nanoparticles, the structure of the magnetic particles was characterized by XRD and the diffractogram is shown in Fig. 5. There are six diffraction peaks: (2 2 0), (3 1 1), (4 0 0), (4 2 2), (5 1 1) and (4 4 0) in the unmodified Fe<sub>3</sub>O<sub>4</sub> particles, which is the standard pattern for crystalline magnetite with spinel structure (Ma et al., 2005). It is clearly seen there is little influence of the dispersant on the structure of magnetic particles, except that the amorphous phase of CS or OCMCS appears as a scattered maximum on the background as seen from Fig. 5(b) and (c).

#### 3.5. Magnetic results

The magnetization of ferromagnetic  $Fe_3O_4$  bulk material is very sensitive to the microstructure of the sample. Superparamagnetism occurs when the particle is small enough that thermal fluctuations (these are of order kT) can overcome the magnetic anisotropy (this is of the order of KV, where K is the anisotropy constant and V is the particle volume). Once this happens the magnetization of the particle is no longer fixed along a certain direction (determined by the easy magnetic direction of the lattice) but appears to be random, in which each particle acts as a big "spin" with suppressed exchange interaction between the particles. The lack of hysteresis is one of the criteria requirements for the identification of the product as superparamagnetic.

Magnetization hysteresis loops at 300 K of unmodified, CS and OCMCS surface-modified Fe<sub>3</sub>O<sub>4</sub> magnetic nanoparticles are shown in Fig. 6. The specific magnetism  $\sigma_s$  is 59.0 emu/g, remanence  $\sigma_r$  is 3.6 emu/g, and the coercivity  $H_c$  is 28.7 Oe for the uncoated Fe<sub>3</sub>O<sub>4</sub> magnetic nanoparticles, while  $\sigma_s$  is 39.1 emu/g,  $\sigma_r$  is 0.5 emu/g and  $H_c$  is 4.5 Oe for CS/Fe<sub>3</sub>O<sub>4</sub>

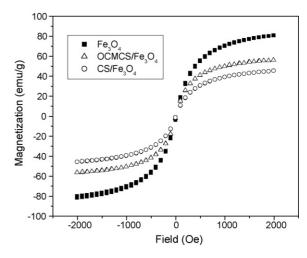


Fig. 6. Magnetization curve for the  $\mathrm{Fe}_3\mathrm{O}_4$  magnetic nanoparticles at room temperature.

magnetic nanoparticles, respectively;  $\sigma_s$  is 51.4 emu/g,  $\sigma_r$  is 0.5 emu/g and  $H_c$  is 5.0 Oe for OCMCS/Fe<sub>3</sub>O<sub>4</sub> magnetic nanoparticles, respectively. The  $\sigma_s$  of the prepared magnetite is high, and the  $\sigma_r$  and  $H_c$  are low. It can be seen from Fig. 6 that  $\sigma_s$ ,  $\sigma_r$ , and  $H_c$  decrease after the particles are coated. The decrease of the magnetization is attributed to the coating polymer on the magnetic nanoparticles. In our experiment the resulted magnetic nanoparticles were immobilized on the wall of a test tube for 10s by applying a magnetic field. The saturation magnetization of CS and OCMCS surface-modified Fe<sub>3</sub>O<sub>4</sub> magnetic nanoparticles was also found to be enough for magnetic separation as a conventional magnet. This method is advantageous over the other methods to get the polymer-modified magnetites, in which, the saturation magnetization was only found to be 16.3 emu/g caused by the low  $Fe_3O_4$  content (24.3%) in the modified Fe<sub>3</sub>O<sub>4</sub> nanoparticles (Ma et al., 2005). And the other magnetic microspheres obtained from copolymerization also contain only small loading of magnetic materials and the process is tedious (Furusawa et al., 1994; Sauzedde et al., 1999).

### 3.6. Stabilization mechanism

A crucial physical property of colloidal dispersions is the tendency of the particle aggregation. Encounters between particles dispersed in liquid media occur frequently, and the stability of the dispersion is determined by the interaction between the particles during this encounter.

To promote a stable dispersion, short-range repulsive forces are required in order to keep each particle discrete and prevent it from amassing as larger and faster settling agglomerates. Steric hindrance is one of the major surface forces playing an important role in stabilizing suspensions. This is accomplished by the coordination of sterically demanding molecules or polymers that act as protective shields on the oxide surface (Bonnemann et al., 2001). The electronic repulsive force results from creation of an electric double layer around the particles, which provides another mechanism to prevent particle agglomeration.

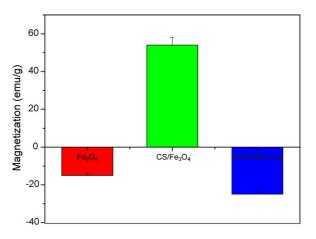


Fig. 7. Zeta potential of colloidal suspension of magnetic nanoparticles (CS/Fe<sub>3</sub>O<sub>4</sub> in pH 4, OCMCS/Fe<sub>3</sub>O<sub>4</sub> in pH 7.4 and Fe<sub>3</sub>O<sub>4</sub> in DI water).

In the present system, the stability of suspension originates from tight binding of chitosan or OCMCS on the surface and charge of the particle, which causes repulsion between the particles. It is well known that because of strong intermolecular hydrogen bonding, chitosan and its derivatives are very susceptible for agglomeration by overcoming the barrier from salvation and electrostatic repulsion. Therefore, a zeta-potential measurement is considered as a key parameter for providing an insight into the charge of the resulting Fe<sub>3</sub>O<sub>4</sub> nanoparticles. The zeta-potential results are shown in Fig. 7. The zeta potential of suspension for unmodified Fe<sub>3</sub>O<sub>4</sub> nanoparticles, CS/Fe<sub>3</sub>O<sub>4</sub> nanoparticles and OCMCS/Fe<sub>3</sub>O<sub>4</sub> nanoparticles is -13.40, +54.20 and -33.45 mV, respectively. The unmodified Fe<sub>3</sub>O<sub>4</sub> magnetic nanoparticles precipitate from their aqueous suspension, which suggests that the zeta potential of -13.40 mVfor unmodified Fe<sub>3</sub>O<sub>4</sub> nanoparticles is not enough to achieve a stable suspension. In pH 4 aqueous CS/Fe<sub>3</sub>O<sub>4</sub> system, the amino groups protonate into cationic charges (NH<sub>4</sub><sup>+</sup>) (Scheme 1). Therefore, CS can adsorb onto Fe<sub>3</sub>O<sub>4</sub> nanoparticles easily by electrostatic attraction, and the amino groups generate positive charge (+54.20 mV) on the Fe<sub>3</sub>O<sub>4</sub> nanoparticles surface. As a result, the intermolecular hydrogen bonding of chitosan is suppressed greatly and the stabilization mechanism for the suspension of CS/Fe<sub>3</sub>O<sub>4</sub> nanoparticles should be electrostatic repulsion. In addition, CS/Fe<sub>3</sub>O<sub>4</sub> nanoparticles with net positive charge can be used to bind with recombinant DNA plasmids to form complex magnetic nanoparticles for targeting genes delivering into cells (Senel et al., 2000; Fang et al., 2001). Also the co-administration of chitosan with drugs and superparamagnetic nanoparticles could enhance the transcellular and paracellular transport of drugs across mucosal epithelium (Chan et al., 2001).

OCMCS has a backbone structure similar to CS, but its *o*-hydroxyl group of each monomer is substituted by a carboxymethyl group through ether bond formation. In pH 7.4 neutral OCMCS/Fe<sub>3</sub>O<sub>4</sub> system, part of carboxylic acid groups will associate into negative charges and the other carboxyl groups keep molecular form (Scheme 1). According to the literature, an aqueous solution of the uncharged poly(vinyl alcohol)

has a strong adhesion to a hydrophilic substance. PVA is known to adsorb on oxide surfaces due to the interaction of hydrogen bonding between the polar groups of the polymer and the surface of the oxide (Lee and Somasundaran, 1989). In our case, OCMCS with functional carboxyl groups is supposed to be chemisorbed on the surface of Fe<sub>3</sub>O<sub>4</sub> by the coordination of "Fe" of Fe<sub>3</sub>O<sub>4</sub> and "O" of carboxyl groups in OCMCS molecules (concluded according to FTIR results). The partly charged carboxyl groups make Fe<sub>3</sub>O<sub>4</sub> magnetic nanoparticles negative (-33.45 mV). Accordingly, the intermolecular hydrogen bonding of OCMCS decreases greatly and the stabilization mechanism for the suspension of OCMCS/Fe<sub>3</sub>O<sub>4</sub> nanoparticles is also the electrostatic repulsion. In addition, using the carboxylic moiety as a binding site, various molecules (e.g., DNA, proteins), and antibodies, could be immobilized on the OCMCS/Fe<sub>3</sub>O<sub>4</sub> nanoparticles for potential specific applications, such as the magnetically targeted drug carriers. Moreover, OCMCS/Fe<sub>3</sub>O<sub>4</sub> nanoparticles are supposed to provide a better association with a hydrophobic drug due to the amphiphilic property of OCMCS. A hydrophobic drug should be located between the OCMCS layer and Fe<sub>3</sub>O<sub>4</sub> nanocore. The research on the targeted release of anticancer drugs from these complex superparamagnetic nanoparticles has been investigated in our lab. Magnetic nanoparticles are generally surface modified with hydrophilic polymers such as albumin (Renshaw et al., 1986), dextran (Molday and Mackenzie, 1982), and starch (Veiga et al., 2000) to disperse them in an aqueous vehicle; however, nanoparticles stabilized in this way have limited applications for drug delivery primarily because of the difficulty of loading these formulations with high doses of therapeutic agents, and especially with water-insoluble drugs. High drug loading on a drug carrier system is necessary to achieve a therapeutic dose of the drug in the target tissue/organ effectively. Rapid dissociation of drug from nanoparticles may induce its premature release in the blood stream, and hence the efficacy of the carrier system to target the drug to the desired tissue/organ and retain it there for a sustained period of time could be significantly reduced. The present procedure is simple and effective process for the preparation of well-dispersed aqueous suspension of magnetite nanoparticles in comparison with the previous protocols reported in the literature.

### 4. Conclusions

The biocompatible chitosan and OCMCS have been proved be an effective dispersant to prepare the well-dispersed suspension of magnetic  $Fe_3O_4$  nanoparticles. The  $Fe_3O_4$  nanoparticles were fully characterized with regard to their morphology, magnetic properties, composition and size distribution by TEM, VSM, FTIR and DLS. The adsorption mechanisms of CS and OCMCS onto magnetic  $Fe_3O_4$  nanoparticles are electrostatic attraction and coordination interaction, respectively. The stabilization mechanism of CS and OCMCS are believed to be electrostatic repulsive. The stable suspension of CS/Fe<sub>3</sub>O<sub>4</sub> magnetic nanoparticles has net positive charge and OCMCS stabilized magnetic  $Fe_3O_4$  nanoparticles have functional carboxyl groups.

### Acknowledgements

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